

# Vitellogenin of *Blattella germanica* (L.) (Dictyoptera, Blattellidae): Nucleotide Sequence of the cDNA and Analysis of the Protein Primary Structure

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**The cloning and sequencing of a cDNA of the vitellogenin gene from the cockroach *Blattella germanica* is reported. It is 5,749 nucleotides long and encodes an amino acid sequence of 1,862 residues (including a putative signal peptide of 17 residues). The vitellogenin sequence includes a long serine-rich stretch between amino acids 322 and 349, and two other stretches between amino acids 1691 and 1740. The vitellogenin of *B. germanica* shows a notable similarity (between 32 and 42%) to those described in other insects, and its alignment shows a high number of motifs conserved in all species, especially in the subdomains I–V. Non-parsimony methods (Neighbor Joining) of phylogenetic analysis of the insect vitellogenin sequences gave a tree showing a topology that is, in general, congruent with the currently accepted insect phylogenetic schemes. Arch. Insect Biochem. Physiol. 45:1–11, 2000. © 2000 Wiley-Liss, Inc.**

**Key words:** insect; Dictyoptera; cockroach; *Blattella germanica*; vitellogenin

## INTRODUCTION

Vitellogenins are proteins that serve as precursors of vitellins, the main egg yolk protein in many oviparous vertebrates and invertebrates. In a great majority of insect species, vitellogenin is synthesized in the fat body, released into the hemolymph, and incorporated into developing oocytes through receptor-mediated endocytosis (Hagedorn et al., 1998; Sappington and Raikhel, 1998). In insects, the endocrine regulation of vitellogenesis has been the object of intensive research, especially in species where the process is ecdysteroid-dependent, like mosquitoes (Hagedorn et al., 1998; Sappington and Raikhel, 1998). Conversely, studies based in less modified species, like cockroaches in which vitellogenesis is juvenile hormone-dependent, are scarcer (see Wyatt and Davey, 1996).

In a previous paper, we reported the isolation and partial sequence of a vitellogenin cDNA on the German cockroach, *Blattella germanica*, and its use as a probe in Northern blot analysis to assess the developmental, tissue and sex specificity of the vitellogenin gene expression in this cockroach (Martín et al., 1998). The same probe was used to study the action of juvenile hormone

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upon vitellogenin synthesis in vivo (Comas et al., 1999), in the frame of our investigations on the functional links between the cycle of such a hormone (Bellés et al., 1987; Maestro et al., 1994) and that of vitellogenin (Martín et al., 1995a,b, 1996; see also Bellés et al., 1999).

Besides these functional studies, we have also been interested in the structural and phylogenetical information that may be derived from vitellogenin primary sequences. Therefore, we have completed the study of the vitellogenin cDNA of *B. germanica*, which, in addition to leading to the characterization of the complete deduced amino acid sequence, has allowed comparison with other species in which the orthologous protein has been reported. Up to now, vitellogenin-based phylogenetical analyses in insects have included species of Hemiptera, Hymenoptera, Coleoptera, Lepidoptera, and Diptera (see Lee et al., 2000a, and references therein). The present work incorporates the “primitive” order Dictyoptera to the analysis, including the complete sequence of vitellogenin of *B. germanica* described herein, and that of *Periplaneta americana* sequenced by Tufail et al., (2000). The comparative analysis will show that the insect vitellogenin molecule seems to contain phylogenetic information.

## MATERIALS AND METHODS

### Insect Rearing and Tissue Collection

Specimens of *B. germanica* were taken from a colony reared in the dark at  $30 \pm 1^\circ\text{C}$  and 60–70% relative humidity. Dissection of fat bodies and extraction of RNA were carried out as described by Martín et al. (1998), although in this case RNA was purified using a Rneasy mini kit (QIAGEN).

### cDNA Cloning and Sequencing

In a previous study (Martín et al., 1998) we obtained a 2,645 nucleotide partial sequence of the vitellogenin cDNA of *B. germanica* corresponding to the 3' region. In order to complete the 5' end of this cDNA we followed the “Rapid amplification of cDNA ends” (RACE) method (Frohman et al., 1988) using the Gibco, BRL kit. An amount of 350 ng of total RNA from fat bodies of females between days 3 and 5 of adult life, were used as starting material. Oligonucleotides to be used as primers were designed on the basis of the 5' end

of the partial sequence and RACE products (see Table 1). PCR conditions were:  $94^\circ\text{C}$  for 2 min;  $94^\circ\text{C}$  for 30 sec, between 60 and  $65^\circ\text{C}$  (depending on the primers) for 30 sec,  $72^\circ\text{C}$  for 2 min, for a total of 30–35 cycles; final extension at  $72^\circ\text{C}$  for 20 min. PCR products were subjected to electrophoresis on 1.2% agarose gel, purified using a QIAquick PCR Purification kit (QIAGEN) and subcloned in Pgem-T Easy vector (Promega). Sequence analysis was directly performed by the dideoxynucleotide chain termination method. Clones were sequenced on both strands using the SP6 and T7 sequencing primers and internal, specific primers, in an automated fluorescence sequencing system ABI (Perkin Elmer, Norwalk, CT).

### Sequence Comparisons and Analysis

Vitellogenin amino acid sequences used for comparison were obtained from literature sources and the GenBank database as follows: silkworm moth, *Bombyx mori* (Yano et al., 1994a,b; D13160); gypsy moth, *Lymantria dispar* (Hiremath and Lehtoma, 1997a,b; V60186); boll weevil, *Anthonomus grandis* (Trewitt et al., 1992; M72980); yellow fever mosquito, *Aedes aegypti* (Chen et al., 1994; U02548); turnip sawfly, *Athalia rosae* (Nose et al., 1997; AB007850); the parasitoid wasp *Pimpla nipponica* (Nose et al., 1997; AF026789); the bean bugs, *Riptortus clavatus* (Hirai et al., 1998; U97277) and *Plautia stali* (Lee et al., 2000a; isoform 1: AB033498, isoform 2: AB033499, isoform 3: AB033500); the cicada *Graptopsaltria nigrofusca* (Lee et al., 2000b; AB026848), and the American cockroach, *P. americana* (Tufail et al., 2000, AB034804). In the case of *P. stali*, only one isoform (Vg-1) was used, given the high percentage of identity between the three vitellogenin

**TABLE 1. Oligonucleotides Used as Primers in 5' RACE to Obtain the Complete cDNA of *Blattella germanica* Vitellogenin\***

| Name  | Sequence (5'→3')       | Position  |
|-------|------------------------|-----------|
| GSP11 | GAAATTCCTGATCTGT       | 3277–3262 |
| GSP12 | GCCCGTGTTTTCTCCCATTT   | 3237–3218 |
| GSP21 | CCTGTCAAGACCTGAAATGTAT | 2991–2970 |
| GSP22 | GTTGTGCGAAAGCTGTGCTGAA | 2964–2944 |
| GSP31 | CCTTGGCTTTCCTTCGCATTT  | 2281–2261 |
| GSP32 | CCACATTTGCAGCACTCTTA   | 2242–2223 |

\*The positions of oligonucleotides are indicated by nucleotide numbers (see Fig. 1).

isoforms known for this species, and to avoid redundancy effects. The software package of the Genetics Computer Group (GCG, version 9.1) of the University of Wisconsin (Devereux et al., 1984) was used for sequence alignments, which were carried out with PILEUP and were not further hand-refined. Percentage similarity and percentage identity between sequences were estimated using the corresponding application of the Multiple Sequence Alignment Editor GeneDoc version 2.5.000 (Nicholas and Nicholas, personal communication, 1997). Phylogenetic analyses were carried out using amino acid sequences, with the Phylogeny Inference Package (Phylip, version 3.57c, Felsenstein, personal communication, 1995). For non-parsimony analyses, we followed the method of Neighbor Joining, and the distances between different vitellogenins were estimated with Kimura's formula and the application Protdist. Parsimony analyses were carried out with the same Package using the Protpars method. Bootstrap analyses were carried out with the application Seqboot in the Phylip package, and the procedure was repeated 100 times.

## RESULTS AND DISCUSSION

### Vitellogenin cDNA and Deduced Amino Acid Sequence

The completion of the partial sequence previously reported (Martín et al., 1998), following the 5' RACE method, yielded a sequence that is 5,749 nucleotides long (Fig. 1). The 5' region begins with a short (24 bp) untranslated region, which is rather constant in size compared to different insect vitellogenin cDNAs (see, for example, Hirai et al., 1998). The translated region begins with the ATG codon corresponding to the initial methionine, and finishes with the stop codon located 5,611 bp downstream. After the stop codon there is an untranslated region of 139 bp, which contains two fused consensus polyadenylation signals (AATAAATAAA), reminiscent of the two complete and successive polyadenylation signals reported by Hiremath and Lehtoma (1997a) in *L. dispar*.

The deduced amino acid sequence has 1,862 amino acids (Fig. 1), and the calculated molecular weight is approximately 212 kDa, which is in fair agreement with the molecular weight of the pre-pro-vitellogenin estimated from electrophoretic data (Martín et al., 1995a). The first 17 amino

acids would correspond to a signal peptide, as suggested by the hydrophobicity analysis (not shown) and by the predictions of the program SignalP V1.1 (Nielsen et al., 1997), and given that they fit the -3, -1 rule of von Heijne (1986). In addition, these 17 first amino acids align well with the signal peptides of other insect vitellogenins (Fig. 2), and show a relatively high percentage of similarity when compared (the highest similarity, 94%, was found between *A. rosae* and *R. clavatus*, and the lowest, 27%, between *B. germanica* and *B. mori*). The similarities in primary structure of these signal peptides (see also Romans et al., 1995) suggest that they are orthologous, and point to a common ancestor signal peptide for insect vitellogenins.

Other remarkable motifs of the amino acid sequence are the polyserine domains. There is a long serine-rich stretch between amino acids 322 and 349, which is 41 amino acids before a consensus RXRR (RPRR) cleavage sequence. There are two shorter serine-rich stretches between amino acids 1691 and 1740. The occurrence of such polyserine domains towards the 3' region of the sequence is only paralleled in the vitellogenins of *A. aegypti* (Chen et al., 1994; Romans et al., 1995) and of *P. americana* (Tufail et al., 2000). Conversely, serine-rich stretches are quite frequent towards the 3' end, around the cleavage site (see, for example, Lee et al., 2000b). The most frequent variant is the occurrence of two stretches flanking the cleavage site, as occurs in *B. mori* (Yano et al., 1994b), *A. aegypti* (Chen et al., 1994; Romans et al., 1995), *A. rosae* (Nose et al., 1997), *R. clavatus* (Hirai et al., 1998), *P. stali* (Lee et al., 2000a), and *P. americana* (Tufail et al., 2000). *G. nigrofusca* has only one serine-rich stretch following the cleavage site (Lee et al., 2000b), and *B. germanica*, in this 3' region, also has only one stretch, but preceding the putative cleavage site. All these variants in distribution of polyserine domains have been schematically represented in Figure 3. *A. grandis* (which is cleaved in the 5' region, as usual) (Trewitt et al., 1992), *L. dispar* (which is cleaved in the 3' region) (Hiremath and Lehtoma, 1997a), and *P. nipponica* (which is not cleaved) (Nose et al., 1997) have no clusters of serine residues at all.





1513 AACTATATGAATTGGGATACATGGAGAGTCTACCGCGATGCTGTGTGCACAAGCAGGTACTTGGTCTGCTCTA  
497 N Y M N W D T W R V Y R D A V S Q A G T W S A L

1585 AATTCAATTCAGCAATTCATCTCTTCTGAAATGGTTGAACCAAAAAGAAGCCTCACATCTCATTACCGTCTTA  
521 N S I Q Q F I S S E M V E P K E A S H L I T V L

1657 CCAGCTGCCGTCTCAGATAAGAACAAGGCGTATCTGCATTTCTTGTGTTGAAATGACAAAGGACCCTGTATTT  
545 P A A V S D K N K A Y L H F L F E M T K D P V F

1729 AAGAACATGACATATGTTAACACCAGCCTCGTTCTAGCATTCTCAGAGGTTATACATCAAGTAGAAATGCAT  
569 K N M T Y V N T S L V L A F S E V I H Q V E M H

1801 CAGGTCAGAGATTGAAAATAAAGTCGGTATATATTCCATACCTGGTCCAAGAATTTCGATGATGCAGTCAAA  
593 Q V R D L K I K S V Y I P Y L V Q E F D D A V K

1873 GAGAATAACAGCATTAAAATTCAGTTATACACCCATGCCCTTGGCGTTACCGGAAACACACACATTCTCCAT  
617 E N N S I K I Q L Y T H A L G V T G N T H I L H

1945 TATCTCCGCCCTATATTATTCAATTGAAGACCATTACTCACCATCAGCGACTCTTCATGGTCCAGTCCTTG  
641 Y L R P Y I I Q L K T I T H H Q R L F M V Q S L

2017 GAAAGAGTGGTAGAACACAACCCTCGAAAGGTTATCGATCTCCTCTTGAGTCTTTATCTGGACCAAAACGAA  
665 E R V V E H N P R K V I D L L L S L Y L D Q N E

2089 CATGCAGATATCAGGGTCGAAGCATTATTCTTGCTAATGAAGGCTGATCCCTCCATCCACGTCCTCAAGATG  
689 H A D I R V E A L F L L M K A D P S I H V L K M

2161 GTTGCAGAACTAACCATACGGAAAGCAACAATCAAGTTCTCTCAGCTTCGCAGTCTGCCATTAAGAGTGTCT  
713 V A E L T H T E S N N Q V L S A S Q S A I K S A

2233 GCAAATGTGGAAGGCGATATATACTCTGAAATGCGAAGGAAAGCCAAGGCCGTGGAACACCTGCTGAGCACA  
737 A N V E G D I Y S E M R R K A K A V E H L L S T

2305 AGGAATATGGATGTATCATACTCCAAGAGTTACTTGTACGGCTATAAGAGCAAAAAATCAATTATGATTCC  
761 R N M D V S Y S K S Y L Y G Y K S K K I N Y D S

2377 CTCTATAATCTAAACTACATTGGAAGTGAGGACAGCATCTATCCTAAGTCCATGCTCTTAAATATCTTTACT  
785 L Y N L N Y I G S E D S I Y P K S M L L N I F T

2449 AACAACTTTGGTAGAATCAACACCCATGTTTCAGAAGGGATATATGGTATCAAGTATGACTGATCTATGGGAA  
809 N N L G R I N T H V Q K G Y M V S S M T D L W E

2521 GCTTTCCACACAATATATAAAAAAGATAATGGATCACCTACTGATCCTAAAACCTTGGTAAAATTTGTTGAA  
833 A F H T I Y K K D N G S P T D P K T L V K F V E

2593 GGCAATCTGAAATACTTTAACATGGGTGTCCAGAAATTTGGGCATTTGACAACACAACATTTCAGTAATGCA  
857 G N L K Y F N M G V Q K F W A F D N T T F S N A

2665 TCTGCAGTAATACAGGAGTTCCTTAAAACCTACAAAAACCTACAAATTTCAACCACACCAAACTCTCCAGC  
881 S A V I Q E F L K T Y K K P T N F N H T K L S S

2737 AGCTCTTCAATTACTCTAACCCCTGCCCTGTGCAATGGGTTTGCCTGCATATTTCAAGATGAACTCACCAAGT  
905 S S S I T L T L P C A M G L P A Y F K M N S P S

2809 TTATGGAAGTATAATGGTGAATTTAGTATTCAGACTGATGCTAAAACCTGATGTACCAATGTCTCTCGAGAAT  
929 L W K Y N G E F S I Q T D A K T D V P M S L E N

2881 TTTATGAACATCACTGGTTCAATTAATCTTATGTTTCAGCCAGATGTACCATGCCCAGCTTGCTTTCAGCACA  
953 F M N I T G S I N L M F S Q M Y H A Q L A F S T

2953 GCTTTCGACAACAAGGAATACATTTTCAGGTCTTGACAGGAAAGTGAAGTTTCATGTTCCAGTAAAATTTCCAA  
977 A F D N K E Y I S G L D R K V E V H V P V K F Q

3025 ATAAATCTGGATTTCAAGAACCACAACGGGTTTCATACGAATCATCCATTATTCACTGATAGAGATTACGAT  
1001 I N L D F K N H N G F I R I I P L F T D R D Y D

Figure 1. (continued).

3097 GTTTTGCAATGGCAGACTATTCCATACACAACCATCCATAACGTACCAGACTTTGAAACAGTCTACATGGAT  
 1025 V L Q W Q T I P Y T T I H N V P D F E T V Y M D

3169 CAACTCTTCAAATTGATCCACGTGCGAAAAACAGCACACTTCGAGAAGAAAATGGGAGAAAACACGGGCATT  
 1049 Q L F K L I H V R K T A H F E K K M G E N T G I

3241 GTATTTAAAGTAAAATATGATACAGATCAGGAATTTCTGGATACAAAAGTGGTTTCTAGATGAATTCAAGGTT  
 1073 V F K V K Y D T D Q E F L D T K W F L D E F K V

3313 CTCCAACCTTTTCACTGGTCTTAATTATGACGTGCCACAAAGGACATCTTCTACAACAACCTGACTGTATAT  
 1097 L Q L F T G L N Y D V P T K D I F Y N N L T V Y

3385 TATGATCATGAAGACACCAAAAACCATGCCGTATCTTTACTGTGACAAAAGAACAATCTAAGTTCTACGAA  
 1121 Y D H E D T K N H A V S F T V T K E Q S K F Y E

3457 ACATTA AACCCAGTAGTTCAACAAAACCTAAAACCTTTCATCTGGAAAAGAAGCAAAAACATCGTAATGTAAAG  
 1145 T L N P V V Q Q N L K L S S G K K Q K H R N V K

3529 AGCCATCGTATAAGGAGAGAATATACCGAGGATGAAAATCCAGCTATTCCAAAAGACAAAACCAACCAACAGC  
 1169 S H R I R R E Y T E D E N P A I P K D K Q P N S

3601 CATCCAGACGTCAAGAATACCTAAGCAAATCAATGGCACTAACTGGAGATGCAACTGCAGTTGTACTTGTAT  
 1193 H P R R Q E Y L S K S M A L T G D A T A V V L D

3673 ATGACTTTGAAGTTTGAGGGGCTGCTGAATCCTACTTTACAACCACCGTATCTCAGCTACATCTCTTGT  
 1217 M T L K F E G P A E S Y F T T T V S H A T S L V

3745 AATGGTAGCTCTAACTACCTACTCTTCTACGATCAACACTACTATGAGGAAAAGAAGAGAAATCAGTTCTGT  
 1241 N G S S N Y L L F Y D Q H Y Y E E K K R N Q F C

3817 CTTTCATGGAGTGTCTACAAACCACAAGTTCCAATCATGAATATTTATAGTGCTTTTGAATTTGATCCAAAT  
 1265 L S W S V Y K P Q V P I M N I Y S A F E F D P N

3889 TCCAAAGTGCATGCCATAATGAACATAGGAAAAGAATGTGAGAATGGAGGAAGTGCTGTAGCAAATATTGAT  
 1289 S K V H A I M N I G K E C E N G G S A V A N I D

3961 ATGCTTAGATTGTGAGAACACCTAGATTACGTAAAGAATTTAACAGTCAGCAAGCTATGCGACCATGAAATG  
 1313 M L R L S E H L D Y V K N L T V S K L C D H E M

4033 CGTACAAAACGTGATCACGTTCTGCCAGCTTGCAGGAATTTCTACTGAGAGAGCCAGTGATCTCAACAGAGTT  
 1337 R T K R D H V L P A C R N S T E R A S D L N R V

4105 CATGTTGATATCAATTATAACTTAAAGCAGCATGAAACATTCAAACGCAGAGTATACAAAGTATATGATTTT  
 1361 H V D I N Y N L K Q H E T F K R R V Y K V Y D F

4177 GTACGTACCCACCTCTATCCACACGTATCAGAGGATGTGATCGTTGATAATCCAGCACAAATTCATATCGGCA  
 1385 V R T H L Y P H V S E D V I V D N P A Q F I S A

4249 AATTTACATTGAAAGACAATACAAGAGCCTTCAACGTGAGCATTGAAACTCCTGTACTTAGTGTTAATGCA  
 1409 N F T L K D N T R A F N V S I E T P V L S V N A

4321 ACTTCGGTTCGACTTCAATCATGGCAAAGTGAATGCTCAGAATGAATCCAAGAAGTTCATTTGCAAAAACGT  
 1433 T S V R L Q S W Q S E M L R M N P R T S F A K R

4393 TTTGCCAAATGGGCTCTACCCCTCTACTATAAACCAACGTGTGTTGTGGATTCTTCTACATCAATACATTC  
 1457 F A K W A L P L Y Y K P T C V V D S S Y I N T F

4465 GACAATTTTACATACAGTGCTCATCATATTGTACAGAACGATGCTTTCTACACTATATTGGATATTCCTCAA  
 1481 D N F T Y S A H H I V Q N D A F Y T I L D I P Q

4537 AAGTTTAATATGAATACTTTAAAGTTGCATTCAAACCAACATCCCAGTTCCAAATATGCAGAGGGAAGTC  
 1505 K F N M E Y F K V A F K P T S P V P N M Q R E V

Figure 1. (continued).

4609      TTAGTGTTCCTGAGAAATGCAAAAATAGAATTGAAACCTAATCAAGGAATGCCAGAAGTTTACGTAGAAGGT  
1529      L V F L R N A K I E L K P N Q G M P E V Y V E G

4681      AAAAGAGTGGATTATAACCACCACCATTCAACTGACTTGAATGTGAGTCAAGACCCGGATTGGTTATGTCTAT  
1553      K R V D Y N H H H S T D L N V S Q D R I G Y V Y

4753      GCCCTTCCAACAAAGGCTGCTCATATTGTATTTCCCTTCATACGAGATCGAAATGTTTTACGATGGCTCCAGG  
1577      A L P T K A A H I V F P S Y E I E M F Y D G S R

4825      ATTATGATACAAGCTTCAAATATGTACCGAAATTTACCAAAGGTCTCTGTGGTAACATGGATGGAGAATTC  
1601      I M I Q A S N M Y R N **F T K G L C G N M D G E F**

4897      GTGAACGACGTGTTGACTCCATGGGGCTGCTACGCCAAAGACATGGCACTCTTTGTAGCATCTTATGCCGAC  
1625      V N D V L T P W G C Y A K D M A L F V A S Y A D

4969      AACAGCAACAGCGAAGTAAGGAAGATCAAGGCTACCCAAAACGAGCAGACATGCGTCCCACAGTTTCATCAA  
1649      N S N S E V R K I K A T Q N E Q T C V P Q F H Q

5041      CCTCTAGTCAGCCATCAAATGAGATTGTCTCAAGTCATTAAGTTAGCAGATACCTCTTCTCCTCTGAATCT  
1673      P L V S H Q M R L S Q V I K L A D T S S S S E S

5113      TCCTCTTCATCAGAATCTCATGAAAACAATTCATCACCATCTTCTGAGTCTCAAGTCAACAAGTCTAAGCGT  
1697      S S S S E S H E N N S S P S S E S Q V N **K S K R**

5185      CAACCAAATTCAGACCAAGATCCAGTTCAGTTCAGCTCCAGCTCCAGTCTGAATCCAATGAATCTGTC  
1721      Q P N S R P R S S S S S S S S S S S S E S N E S V

5257      CTTGCCAAGAAAATCATTAAACAACCAAATGGACCAAAGCCTACTTAATACCATCACAGTCTCCTATGACG  
1745      L A K K I I N N Q I G P K P T L I P S Q S P M T

5329      TCCGATGACAAATGCATGACACAACAACCAAGACACACTTATTATGAAAACCAATCTGTGTGAGTGA  
1769      S D D K C M T Q Q P R H T Y Y E N Q F C V S E K

5401      CCCCTGGACACATGCATGCCCTTATTTGCCATGCAACTGAATCCTATACAATAGATGTAATTTTTACTGT  
1793      P L D T C M P L I C H A T E S Y T I D V N F Y C

5473      GTTCCACTAGGTCCTGCAGCTAATCATTACATGAAACTTGTCAAGAAAGGCATTCTCCCAGATTTAAGTCGG  
1817      V P L G P A A N H Y M K L V K K G I L P D L S R

5545      AACAGGAATGGTAAGAGGGTAGTTCTTCTCTGTGAAATCCCAATACAATGCGAACCTGTTTTAAACTGAAAT  
1841      N R N G K R V V L P V E I P I Q C E P V L N \*

5617      CCGATCTTGTTAATGGACTTAATTTAAAAAATCTGACTTATGAAGAAAACCTGGAATATTTTCAAAAATGA  
5689      ACTATTTATGTATTT**AATAAATAA**GAATGATAGCAACTTGAAAAAAAAAAAAAAAAAAAAA

Fig. 1. Nucleotide sequence and deduced amino acid sequence of vitellogenin cDNA of *B. germanica* (GenBank accession number: AJ005115). The putative signal peptide is indicated by a single underline. Clusters of serine residues are indicated by double underline. Boldfaced letters

indicate putative asparagine-linked glycosylation sites. Putative cleavage sites (having the consensus sequence RXXR) for the proteases of the subtilisin family are framed. Asterisk means stop codon. Italic boldfaced letters show the double polyadenylation signal.

In addition, the analysis of the amino acid sequence reveals six putative cleavage sites (having the consensus sequence RXXR) for the proteases of the subtilisin family (Barr, 1991) at positions 119–122, 283–286, 391–394, 505–508, 1171–1174, and 1337–1340. Finally, putative asparagine-linked glycosylation sites are present at positions 247, 278, 570, 575, 618, 874, 879, 898,

955, 1116, 1241, 1325, 1349, 1409, 1420, 1431, 1482, 1566, 1611, 1705, 1716, and 1741.

**Comparison With Vitellogenin Sequences of Other Insect Species**

Comparison of the amino acid sequence of *B. germanica* (Fig. 1) with those of other insect species revealed that the percentage of similar-

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Blattella      : MTTNALCCLLVSAASA----
Periplaneta   : -MKGFCCLLVAGVTS----
Graptopsaltria : -MWAPLYMCLLVAGAIA----
Plautia1      : MLWSSAE--LLAFACLAAA--
Plautia2      : MNWTLVA--LLTFVGLAAA--
Plautia3      : -MWAPFT--LFVVAFRTLASA
Riptortus     : -MWSPVVLCLLVGLASA----
Anthonomus    : -MWSTVALCCLLVGLSYVSSS-
Pimpla        : -MWCPLFLVLLAGAATA----
Athalia       : -MWSPLVLCCLLVGIASA----
Aedes         : -MLAKLFLALAGLTAAYQY-
Anopheles     : -MIAKLLLLTLVGLCT-----
Lymantria     : -MRLLLISAFIAVSS-----
Bombyx        : -MKLFVLAATIAAVSS-----
Locusta-A     : -MNAVTVLGGL??????????
Locusta-B     : -MWALVSGLL????????????

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Fig. 2. Alignment of the signal peptide of vitellogenins from the following insects: *Blattella germanica*, *Periplaneta americana*, *Graptopsaltria nigrofuscata*, *Plautia stali* (isoforms 1, 2, and 3), *Riptortus clavatus*, *Anthonomus grandis*, *Pimpla nipponica*, *Athalia rosae*, *Aedes aegypti*, *Anopheles gambiae*, *Lymantria dispar*, *Bombyx mori* and *Locusta migratoria* (isoforms A and B). Database accession numbers for most of the species are indicated in Materials and Methods. Those of the partial sequences corresponding to *L. migratoria* are: M17333 (isoform A) and M17334 (isoforms B) (Locke et al., 1987). The signal peptide sequence of *A. gambiae* is reported in Romans et al. (1995).

ity decreases approximately in parallel to the decrease of phylogenetic relationships (*P. americana*: 42%, *G. nigrofuscata*: 39%, *R. clavatus*: 39%, *P. stali* (isoform 1): 37%, *P. nipponica*: 37%, *A. grandis*: 37%, *A. aegypti*: 36%, *B. mori*: 34%, and *L. dispar*: 32%). The exception is the hymenop-

teran *A. rosae*, which shows a remarkably high (41%) percentage of similarity.

An alignment of the vitellogenin sequences (excluding the signal peptide) of the 11 insect species mentioned above was carried out with the Pileup program from the GCG package (not shown), and revealed numerous conserved motifs (like the tetrapeptide GLCG in position 1615–1618) typical of vitellogenin sequences. The subdomains I–V, described as regions of relatively high amino acid conservation (Chen et al., 1997; Sappington and Raikhel, 1998), were also readily identifiable in the vitellogenin of *B. germanica* within the alignment (subdomain I: between amino acid 22 and 303; II: 442–826; III: 855–1034; IV: 1470–1646; V: 1788–1862).

Finally, we carried out a phylogenetic analysis of the vitellogenin sequences of the 11 available insect species following parsimony and non-parsimony approaches. Parsimony analyses were performed with the Protpars program from the Phylip package, and using the above-mentioned Pileup alignment. The topology of the resulting tree (not shown) presented serious incongruencies with respect to the currently accepted insect phylogenetic schemes (see, for example, Kukulová-Peck, 1991; Friedrich and Tautz, 1995, 1997). For example, the lepidopterans *L. dispar* and *B. mori* appeared as the sister group of all other orders, whereas the dipteran *A. aegypti* appeared in a very internal node. This could mean that the evolution of insect vitellogenins has not been strictly parsimonious.

Non-parsimony analyses were carried out fol-



Fig. 3. Schematic representation of the variants in distribution of polyserine domains in the vitellogenins of different insects. See the binomial nomenclature of all species in the legend of Figure 2. In the case of *Plautia stali*, isoform 1 has been represented. The serine-rich stretches are indicated as black segments, their length being proportional to the length of the stretch. The arrows and the narrow lines indicate the putative or demonstrated cleavage site.



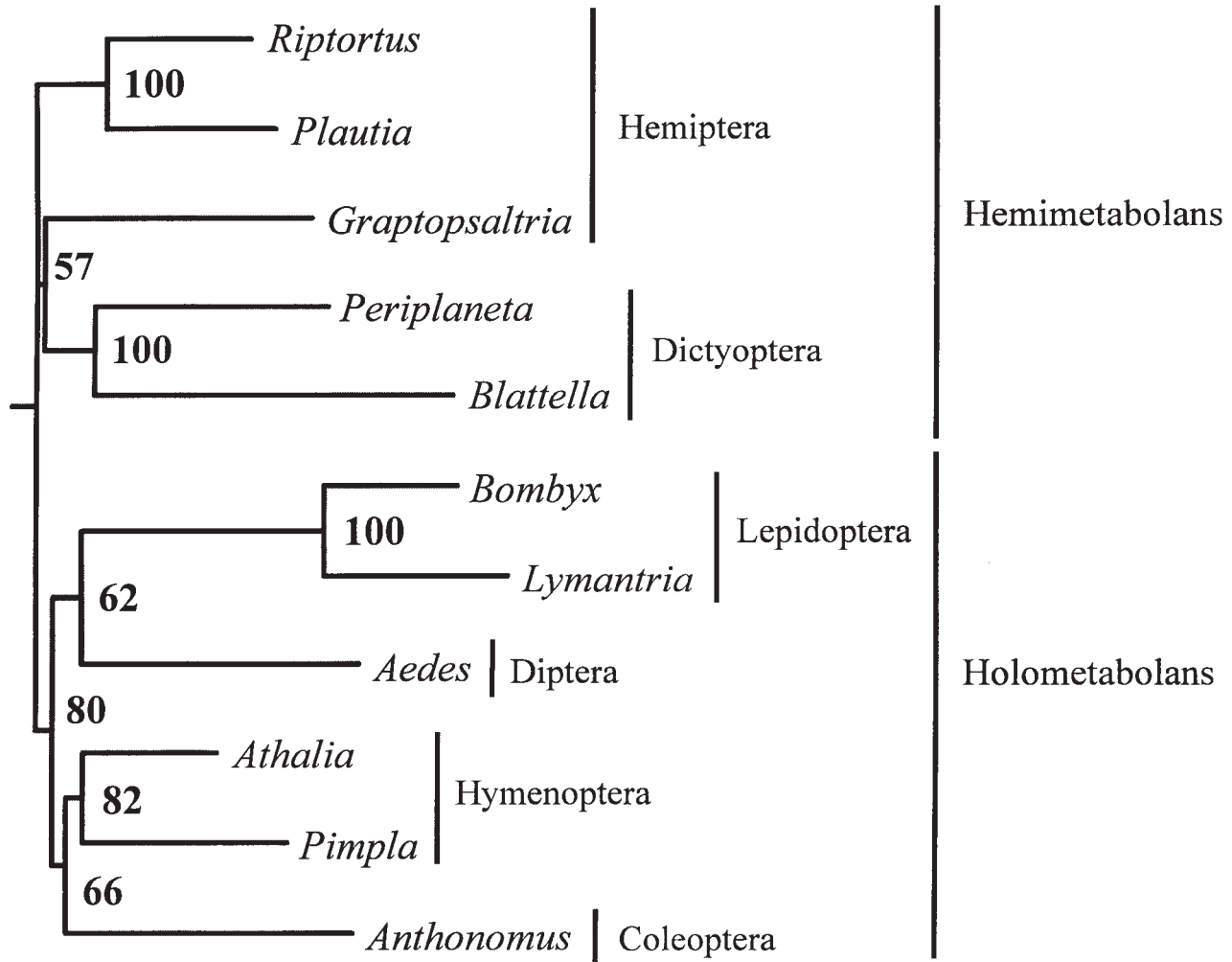


Fig. 4. Phylogenetic tree for the entire sequence of vitellogenin (less the signal peptide) in insects. It was obtained with the method of Neighbor Joining, using the distance of

Kimura. Numbers in the nodes correspond to bootstrap values in 100 replicates. See the binomial nomenclature of all species in the legend of Figure 2.

lowing the method of Neighbor Joining, from the Phylip package, using the distance of Kimura and the entire sequences of vitellogenin (less the signal peptide). The most external nodes of the tree obtained (Fig. 4) separate holometabolans from hemimetabolans. The more internal ones group in a quite coherent manner the different orders (for example dipterans + lepidopterans), and the different species of the same order (for example *B. mori* + *L. dispar*). The sole readily apparent incongruence is the position of the cicada *G. nigrofusca* clustering with the two dictyopterans (although this node was the one having the lowest bootstrap index) and not with the other hemipterans. The same analysis was carried with each one of the five subdomains of

Chen et al. (1997) (see also Sappington and Raikhel, 1998) but gave poorer results (not shown), the orders being incongruently distributed in most cases. Only the analysis based on subdomain II resulted in a relatively coherent topology, although in this case *G. nigrofusca* clustered with the dipterans and lepidopterans. Again, this node was the one showing the lowest bootstrap index (47).

The results of the phylogenetic analysis suggest that the entire vitellogenin sequence should be used when searching phylogenetic inferences. In addition, the incongruent results emerging from parsimony methods of analysis suggest that the molecular evolution of insect vitellogenins has not been strictly parsimonious. However, this would not be exceedingly surprising taking into

account the extreme diversity of reproductive strategies occurring in insects (Engelmann, 1970; Bellés, 1998), which may have resulted in a great diversity of selective pressures acting upon such a molecule.

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